REVIEW



A Short Review: Bioactivity of Fermented Rice Bran

Department of Food Technology, Universitas Bakrie, Kawasan Epicentrum, Jalan HR Rasuna Said Kav C.22, Jakarta 12920, INDONESIA

Abstract: Rice bran is a by-product of the rice milling process, which refers to the processing of brown rice into polished rice. It contains a considerable amount of functional bioactive compounds. However, the utilization of these compounds is limited and calls for an effort to ferment rice bran. One of the methods that can significantly increase the added value of rice bran as well as its bioactivity is the solid-state fermentation. It can also be one of the strategies that help in the production of rice bran as a functional ingredient with higher bioactivity for health promotion.

Key words: bioactive compounds, bioactivity, fermentation, fermented rice bran, solid-state fermentation

1 Introduction

Rice is one of the most important food commodities and is the second-largest agricultural commodity cultivated worldwide¹⁾. World paddy production reached 769.9 million tons in 2018, while the milled rice production, mostly in Asian countries, was around 510.6 million tons²⁾. In line with the population growth, the demand for rice is estimated to continue rising in the next few decades, followed by an increase in the number of by-products from the rice milling process. From 1994 to 2018, Indonesia has been the third-largest rice producer in the world. In 2019, the production of milled dry paddy rice in Indonesia was 54.60 million tons³⁾. Generally, the amount of rice bran is 10% of brown rice by milling process; this implies that 5.46 million tons of rice bran were produced through rice milling. The high amounts of rice bran produced can be utilised optimally with the proper processing methods.

The rice milling process produces rice as the main product and rice husk and bran as the by-products (Fig. 1). The rice grain or paddy has an outer-part(husk) to protect or cover the inner-part of rice. The inner-part of rice consists of a bran layer and endosperm. Brown rice comprises the outer layers of pericarp, seed-coat and nucellus, the germ or embryo and the endosperm. Currently, rice bran is being widely studied owing to its potential as a functional ingredient in food products. It has a high nutritional content, including vitamins, minerals and essential amino acids (tryptophan, histidine, cysteine and arginine)⁴⁾. Moreover, it contains bioactive compounds such as γ -oryzanol, α -tocopherol, tocotrienols, polyphenols, phy-



Fig. 1 Paddy structure.

tosterols, carotenoids, ferulic acid and $phytosterols^{5-8)}$. These compounds serve as antioxidants and have anti-atherosclerotic, anti-diabetic, anti-cancer and anti-inflammatory properties^{9,10)}.

There are still several obstacles in the efforts made to develop rice bran as a functional ingredient, such as the lack of public awareness about its health benefits, a few downstream industries' interest in developing rice bran and the quality of rice bran being sub-standard. In addition, rice bran contains anti-nutritional substances such as trypsin inhibitor, hemagglutinin-lecithin and phytic acid, and its instability during storage is another reason that hinders its use as a food ingredient^{11, 12}.

One of the methods that can be employed to increase

*Correspondence to: Ardiansyah, Department of Food Technology, Universitas Bakrie, Kawasan Epicentrum, Jalan HR Rasuna Said Kav C.22, Jakarta 12920, INDONESIA

E-mail: ardiansyah.michwan@bakrie.ac.id ORCID ID: https://orcid.org/0000-0003-2110-344X Accepted July 23, 2021 (received for review June 9, 2021) Journal of Oleo Science ISSN 1345-8957 print / ISSN 1347-3352 online

http://www.jstage.jst.go.jp/browse/jos/ http://mc.manusriptcentral.com/jjocs



the added value of rice bran for further use in product development fermentation, especially solid-state fermentation (SSF). SSF is a fermentation technique that can escalate the bioactive compounds in food, such as the phenolic content, thereby contributing to antioxidant activity¹³⁻¹⁵⁾. Mold, yeast and bacteria are the types of microorganisms that are often used in the SSF method. Mold species are suitable for use in SSF because they are capable of producing enzymes like amylase, pectinase, xylanase, cellulase, chitinase, protease, lipase and β-galactosidase¹⁴⁾. Therefore, SSF helps to increase the bioactive compounds of plant materials. In addition, fermentation using the SSF method can improve the sensory profile of the rice bran¹³⁾.

SSF is currently used in a wide range of applications from traditional applications such as tempe fermentation in Indonesia. Recently, it has been used to develop new food ingredients like bioactive compounds. It was also a new trend regarding bioethanol and biodiesel production as new energy sources that used agricultural by-products as substrate with microbe as starter. SSF has the following advantages: (1) It is a promising method for obtaining a high yield of bioactive compounds that require less water for microorganisms' growth; (2) It can convert cheap agroindustrial by-products into products that contain various valuable compounds; (3) It can minimise microbe contaminants; (4) It is a simple processing technique and is more practical; (5) It involves high productivity; and (6) Its media conditions resemble the actual habitat¹⁴⁾.

This review discusses the bioactivity of fermented rice bran, which has the potential to be developed as a food ingredient. The discussion begins by defining fermented rice bran, the active compounds contained therein, both volatile and non-volatile compounds and *in vivo* examination to show fermented rice bran's bioactivities.

2 Fermentation Can Increase the Active Compounds of Rice Bran

Fermentation has been widely used to improve the quality of food. It is popular thanks to the extended storage period that it offers, which improves taste and increases the bioactive compounds and protein content of the food item^{11, 16, 17}. Several studies have shown that fermented rice bran has a higher content of bioactive compounds and better functional properties than non-fermented rice bran. The increase in phenolic compounds after the fermentation process occurs due to the breakdown of complex compounds that bind to lignocellulose or polysaccharides^{18, 19}. During the fermentation process, the microbes as starters produce enzymes that can hydrolyse complex compounds in the bound form into compounds in the free form¹⁹. Besides being able to increase the bioactive compounds and antioxidant activity, the fermentation process can

improve the sensory profile of rice bran. Cempo Ireng and Inpari 30 fermented rice bran with *Rhizopus oligosporus* for 72 hours are more acceptable by the 75 naïve panellists than the benchmarks(non-fermented rice bran) because the sample had a dominant liking of both aroma and taste²⁰.

Compared to non-fermented rice bran, rice bran fermented with *Rhizopus oryzae* for 24 hours can increase the total phenolic compounds (TPC) up to five times and inhibit 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radicals up to $87\%^{21}$. In addition, the TPC in fermented rice bran using *Rhizopus oryzae* for 120 hours increased from 2.4 mg/g to 5.1 mg/g¹³⁾.

The fermentation process aims to increase the nutritional content and bioactive compounds in rice bran. Several studies have been conducted to determine the effects of fermentation on the nutritional content, antioxidant activity and bioactive compounds in rice bran. The fermentation process is carried out using molds, such as *Monascus pilosus*¹¹⁾, *Monascus purpureus*^{22, 23)}, *Rhizopus oryzae*^{19, 24, 25)}, *Aspergillus oryzae*²⁵⁾, *Rhizopus oligosporus*^{20, 23, 26–28)}, lactic acid bacteria (*Pediococcus acidilactici*, *Lactococcus lactis* and *Pediococcus pentoseous*)²⁹⁾ and mushrooms (*Pleurotus sapidus*)³⁰⁾.

SSF is a fermentation method effective for increasing antioxidant activity and TPC in food with solid media³¹⁾. The mold *Rhizopus* sp. and *Aspergillus* sp. are proper cultures that can be used in the SSF method since they do not produce toxic compounds during fermentation³²⁾. Some of the research results regarding fermented rice bran are shown in **Table 1**. Some of the fermentation processes of rice bran were undertaken using cultures like *Rhizopus* oryzae, *Rhizopus* oligosporus, *Lactobacillus* plantarum, *Monascus* pilosus, *Aspergillus* kawachii, lactic acid bacteria and *Saccharomyces* boulardii.

Using lactic acid bacteria starter and *Rhizopus oryzae* in fermented rice bran produces lactic acid³³⁾. However, the use of *Rhizopus oryzae* is more desirable owing to the easy separation between the mold biomass and the substrate. Several studies have reported that the fermentation process in rice bran can increase the bioactive compounds. This occurs due to the production of extracellular enzymes which affect the increase in the bioactive compounds in the substrate¹⁷⁾. Furthermore, the microorganisms used as starters in the fermentation process will synthesise compounds and activate metabolic pathways to adapt to the substrate³⁴⁾.

Fermented rice bran using *Rhizopus oryzae* increases the ash, fibre, protein, amino acids, phospholipids and the total phenolic content. The increase in ash in fermented rice bran is due to the synthesis of mycelia³⁵⁾. Another study showed that the increase in the fibre content is due to the intrinsic production of chitin¹⁷⁾ which is one of the compounds of the hyphal cell wall on *Rhizopus oryzae*³⁶⁾.

Fermented Rice Bran

Rice cultivars	Fermentation time (h)	Starter (microbes) Results		References
Neptune, Wells, Red Wells	24	Saccharomyces boulardii	Increase in bioactive compounds and decrease in lymphocyte B cell	5)
Polished rice (Yongin)	240	Monascus pilosus	Increase in total flavonoid	11)
Polished rice (IRGA)	96	Rhizopus oryzae	 Increase in phospholipid and unsaturated fatty acid Reduce total lipid and saturated fat 	15)
Polished rice (IRGA)	96	Rhizopus oryzae	 Increase in ash content, dietary fiber, protein, and amino acid. Decrease in water content, lipid, and phytic acid 	17)
Polished rice (IRGA)	120	Rhizopus oryzae	Increase in total phenolic content	19)
Inpari 30 and Cempo Ireng	48, 72, and 96	Rhizopus oligosporus, Rhizopus oryzae and mixtureIncrease in total phenolic content and antioxidant activity		20)
Polished rice (IRGA)	24	Rhizopus oryzae	 Increase in ash content, dietary fiber, and protein content Decrease in lipid content 	21)
Polished rice	288	<i>Rhizopus oligosporus</i> (strain F0020), <i>Monascus</i> <i>purpureus</i> (strain F0061) and mixture	Increase in total phenolic content and antioxidant activity	23)
Inpari 30	72	Rhizopus oligosporus	Increase in total phenolic content and antioxidant activity	28)
Polished rice	96	<i>Rhizopus oryzae</i> (CCT 7506)	Increase in dietary fiber, ash content, protein, lipid and total phenolic content	35)
Polished rice	44 24	 a. Aspergillus kawachii b. Lactic acid bacteria (Lactobacillus brevis, Lactobacillus rhamnosus, Enterococcus faecium) 	Increase in dietary fiber, lipid content, and total phenolic content	38)

Rhizopus oryzae also produces phytase that hydrolyses complex proteins leading to an increase in dissolved protein³⁶⁾. Phytic acid degradation by *Rhizopus oligosporus* can increase the content of minerals, such as iron, magnesium and zinc, which are bound to phytates³⁷⁾. The presence of phytase also accelerates the hydrolysis of phytic acid, thereby affecting the decrease in its content¹⁷⁾. This further leads to the enhancement of phenolic compounds because of the degradation of lignocellulose by the enzymes in *Rhizopus oligosporus*²⁰⁾. The increase in phenolic compounds also occurs in fermented rice bran with the use of *Rhizopus oligosporus* and *Aspergillus kawachii* combined with lactic acid bacteria (*Lactobacillus brevis*, *Lactobacillus rhamnosus*, *Enterococcus*

faecium)^{28, 38)}. Several studies have also shown a decrease in moisture, lipid and phytic acid levels. An increase in the temperature during fermentation causes a decrease in water content¹⁷⁾. The lipids are used by fungi to form mycelia, resulting in decreased lipid levels³⁶⁾.

Rhizopus oligosporus is the main mold and is widely used as a starter in making tempe in Indonesia. Studies on fermenting rice bran with *Rhizopus oligosporus* have been reported by several researchers^{23, 28)}. The research revealed a similar increase in phenolic compounds in fermented rice bran, which occurs due to the activity of hydrolytic enzymes of fungi such as β -glucosidase²⁵⁾; these enzymes can increase the hydroxyl molecule, which can result in a rise in the amount of free phenolic compounds

in the rice bran³⁹⁾. The use of mixed cultures (*Rhizopus* oligosporus, *Rhizopus* oryzae and other mixtures) in the fermenting of rice bran has also been reported. Fermenting rice bran using the *Rhizopus* oligosporus starter produced the highest DPPH radical scavenging activity, with a fermentation time of 96 hours on Inpari 30 cultivar²⁰⁾.

3 Volatile Compounds of Rice Bran

Volatile compounds play a role in the formation of aroma in food products. They give either a pleasant or unpleasant aroma. They can form along the food chain, starting after harvesting, and occur throughout post-harvest handling, distribution and storage. The heating treatment can result in the loss of volatile compounds due to their volatility. Several previous studies have discussed the volatile compounds found in rice and rice bran. The compounds that have been identified include alcohol, alkanes, ketones and aldehydes as shown in **Table 2**.

Our group has identified and compared the volatile compounds of fermented and unfermented Inpari 30 and Cempo Ireng rice bran²⁰⁾. The extraction of volatile compounds was carried out using the headspace solid-phase microextraction method and analysed using the Gas Chromatography-Mass Spectroscopy (GC-MS) instrument. The analysis showed that there were 57 volatile compounds, including alcohol (23%), aldehyde (19%), acid (11%), ketones and esters (9%), ester (7%), terpenes and benzene (5%), furans and lactones (3%) and pyridine, as well as pyrazine and thiazole (2%). The formation of volatile compounds happens as a result of the lipid degradation reaction during fermentation and the Maillard reaction due to the sterilisation process. Most of the volatile compounds in rice bran under fermentation treatment originate from the lipid degradation reactions. In Inpari 30 cultivar, they were 3-methyl-3-butenol, 2,3-butandienol, benzylalcohol, glycerine, methyl hexadecanoate, (E)-9-methyl octadecanoate, (Z,Z)-9,12-methyl octadecadienoate, 1R- α -pinene, caryophyllene, 2-methoxyphenol and 3-methyl pyridine. However, in fermented Cempo Ireng rice bran cultivar, the volatile compounds were dominated by 4-methyl-3-pentenol, benzylalcohol, glycerine, methyl hexadecanoate, ethylbenzene and caryophyllene.

The fermentation resulted in the rice bran having sweet, creamy, fatty, smoky and green aromas. Smoky and green aromas are thought to trigger the rancid aroma in rice bran due to the lipid degradation by enzymes during the fermentation process. In non-fermented rice bran, the volatile compounds are mostly formed by the Maillard reaction which is influenced by the sterilisation process. In the fermented Inpari 30 rice bran cultivar, the volatile compounds that are thought to be dominant include 2-furanmethanol, nonanal, methyl tetradecanoate, phenol and 2-methoxy-4-vinylphenol. Meanwhile, 2-furanmethanol, hexanal, naphthalene, $1R-\alpha$ -pinene and 2-methoxy-4-vinylphenol are the volatile compounds in fermented Cempo Ireng rice bran. These compounds are thought to form a burnt, nutty and fatty smell. In the non-fermented rice bran, hexanal and nonanal compounds were also found, obtained from the lipid degradation reaction which is thought to produce a rancid aroma. Apart from that, other research⁴³⁾ state that 4-vinylphenol is the main component that causes the unpleasant odour. The difference in the volatile compounds of several rice varieties, such as Inpari 30 and Cempo Ireng, emerges from the differences in the varieties, planting locations, nutritional contents and bioactivities in the rice bran.

Group volatile compounds	Compounds
Alcohols	<i>n</i> -Methanol, <i>n</i> -Ethanol, <i>n</i> -Butanol, <i>n</i> -Propanol, <i>n</i> -Pentanol, <i>n</i> -Hexanol, 1-Octen-3-ol, <i>n</i> -Nonanol, <i>n</i> -Octanol, Benzyl alcohol, 3-methyl-1-butanol, Linalooloxide, <i>n</i> -Heptanol, Linalool, Furfuryl alcohol, 2-phenethyl alcohol, 1-Dodecanol
Alkanes	<i>n</i> -Heptadecane, <i>n</i> -Oktadecane, <i>n</i> -Heneicosane, <i>n</i> -Tetracosane, <i>n</i> -Tetradecane, <i>n</i> -Decane, <i>n</i> -Undecane, <i>n</i> -Dodecane, <i>n</i> -Tridecane, <i>n</i> -Pentadecane, <i>n</i> -Hexadecane, <i>n</i> -Nonadecane, <i>n</i> -Eicosane, <i>n</i> -Dococane, <i>n</i> -Cyclohexane, <i>n</i> -Pentacosane, <i>n</i> -Hexacosane, <i>n</i> -Heptacosane, <i>n</i> -Hentriacontane
Ketones	 6-Methyl-5-hepten-2-one, 2-Decanone, 6,10,14-Trimetil-2 Pentadecanone, Aceton, 3-Pentena-2-on, 2-Heptanone, 6-Methyl-3,5-heptadien-2-one, 3-Octanone, 2-Octanone, 2-Nonanone, 3-Octena-2-on, Isophrone, 2-Undecanone, 4-Methylacetophenone, Geranylacetone, β-Ionone, 2-Heptadecanone, 2-Oktadecanone, 6,10-Dimethyl-5,9-Undecadienane-2-on
Aldehydes	3-Methyl-1-butanal, <i>n</i> -Pentanal, <i>n</i> -Hexsanal, <i>n</i> -Nonanal, Benzaldehyde, Ethanal, Propanal, Isobutanal, Isopentanal, <i>n</i> -Heptanal, <i>trans</i> -2-Hexanal, <i>n</i> -Octanal, <i>trans</i> -2-Heptenal, <i>trans</i> -2-Octenal, 3-Furfural, 2-Furfural, <i>n</i> -Decanal, <i>trans</i> -2-Nonenal, 5-Methylfurfural, Phenylacetaldehyde, <i>trans</i> -2- Decenal, Deca-2,4-diena-1-al, 2-Phenyl-2-butenal

 Table 2
 Volatile compounds in polished (white) rice cultivar.

References: 40-42)

Almost the same components—aldehydes, alcohols, alkenes and ketones—were also found in red rice and black rice bran cultivars (**Table 3**). The β -ocimene component in red rice bran is known to have an aroma like damp clothes⁴⁴⁾. Meanwhile, myristic acid compounds are known to carry a waxy and fatty aroma⁴⁵⁾, and the compounds 6,10,14 trimethyl-2-pentadecane are known to have an aroma of oil, celery and wood⁴⁶⁾.

The guaiacol component is reported to be the main cause of the aroma of black rice. In black rice bran, the nonanal compound is known to have a citrusy, green and fatty aroma⁴⁷⁾. Meanwhile, caproic acid and pelargonic acid have a fatty, cheesy and waxy aroma^{48,49)}.

4 Non-volatile Compounds of Rice Bran

Non-volatile compounds of rice bran have been report-

ed⁵⁰⁾ on the Calrose. Dixiebelle and Neptune rice varieties that grow in the south-eastern part of California, United States. In this study, 465 metabolites were found, consisting of amino acids(126), carbohydrates(35), vitamins and cofactors(28), energy(11), lipids(137), nucleotides(40), peptides (28), secondary metabolites (55) and xenobiotics (8). The antioxidant compounds of rice bran include amino acids (4-guanidinobutanoate and taurine), vitamins and cofactors (tocopherols and tocotrienols) and secondary metabolites (ergothioneine and guinates)⁵⁰⁾ which can potentially be used as antioxidants to inhibit chronic diseases and infections. Other studies have shown an increase in metabolite diversification in fermented rice bran when using Saccharomyces boulardii. The use of specific rice varieties and fermentation treatment on rice bran affects the amount and type of active compounds $produced^{5, 17, 51}$. Several studies related to the identification of non-volatile compounds in rice bran are presented in Table 4.

 Table 3
 Volatile compounds in red and black rice bran cultivars.

Rice cultivars	Volatile compounds					
Red rice	Myristic acid, Nonanal, (E)-β-ocimene, 6, 10, 14 Trimethyl-2-pentadecanone, Hexanal, Octanal, 2-Methylnaphtelene, Benzaldehyde, 2-Heptanon, 1-Octen-3-ol, <i>n</i> -Hexanol, Heptanal, <i>n</i> -Heptanol, 2-Pentyl-furan, (2E)-octenal, <i>n</i> -octanol, (2E)-nonenal, Decanal, (2E)-decanal, (2E,4E)-decadienal, (2E)-undecanal, Geranylacetone, Methyl linoleat, Methyl oleat					
Black rice	Myristic acid, Nonanal, Caproic acid, pentadecanal, Pelargonic acid, Guaiacol, Hexanal, Isovaleric acid, <i>n</i> -Hexanol, Enantic acid, p-menthan-3-one, Caprilic acid, 2-Hydroxy-6-methylbenzaldehyde, (2E,4E)- nonadienal, <i>n</i> -Tetradecane, Geranylacetone, <i>n</i> -Pentadecane, <i>n</i> -Hexadecane, Tetradecanal, <i>n</i> -Heptadecane, Methyl hexadecanoate, Palmitic acid, Ethyl hexadecanoate					

Reference: 43)

 Table 4
 Non-volatile compounds of rice bran.

Rice cultivars	Non-volatile compounds	References
Neptune, Wells, and Red Wells	Galactose, Palmitic acid, α-Linoleic acid, Disaccharide, Xylitol, Glucitol, Alanine, Phosphoric acid, D-Fructose, Sorbitol, and 1, 2, 3-Propane tricarboxylate	5)
Calrose, Dixiebelle, and Neptune	 Amino acid: tryptophan, leusine, isoleusine, valine, fenilalanine, tirosine, poliamine, guanidine, and acetamido Vitamin and cofactor: ascorbate, aldarate, glucorate, and teoronate Metabolite seconder: quinate, ergotionein, benzoate, piperidine, tartrate, and α-amirine 	50)
Basmati 217, Basmati 370, Gambiaka, Shwetasoke, DM 16, Kaho Gaew, Dorado, Sawa Mahsuli, Chennula, Njavara, Calrose, RBT 300, Jasmine 85, IAC 600, LTH, SHZ-2, and Rang Jey	Amino acid (Aspartate, Serine); Fatty acid; Oxylipins; Phospholipid; Carbohydrate; Tocopherol; Nucleotide; and Benzoate	51)
RiceberrySterol: 24-Methylene-ergosta-5-en-3B-ol, 24-Methylene-ergo ol, fucosterol, gramisterol, campesterol, stigmasterol, and β-si Triterpenoid: Cycloeucalenol. 24-methylenecycloartanol		52)
Inpari 30 and Cempo Ireng	Adenosine	57)

The compounds in Thai Riceberry bran are classified into sterol and terpenoid groups⁵²⁾. The identified sterol groups include fecosterol, gramisterol, stigmasterol and campesterol. Fecosterol, commonly found in seaweed, is also found in rice bran and functions as an antioxidant compound⁵³⁾. Gramisterol can be extracted from the rice varieties of Riceberry and exhibits anti-cancer effects against acute myeloid leukemia⁵⁴⁾, whereas stigmasterol has the ability to bind chondrocyte membranes and has anti-inflammatory and anti-catabolic effects⁵⁵⁾. Campesterol is a type of phytosterol that is widely available in plants, and phytosterol decreases the levels of low-density lipoprotein (LDL) cholesterol⁵⁶⁾.

Non-volatile compounds in fermented rice bran using Rhizopus oligosporus in Inpari 30 and Cempo Ireng rice bran varieties have also been reported⁵⁷⁾. Fermentation was carried out for 72 hours at room temperature of 30° C with SSF. The analysis of non-volatile compounds was carried out using the ultra-performance liquid chromatography-tandem mass spectrometer (UPLC-MS/MS) and electrospray ionisation (ESI) mass spectrometry in positive ion mode. There were 72 compounds identified and categorised into secondary metabolites (50%), lipids (22%), amino acids(11%), vitamins and cofactors(10%), peptides (4%), nucleotides (1%) and carbohydrates (1%). Fermentation in Inpari 30 and Cempo Ireng rice bran produces new compounds from the metabolism of tyrosine, phenylalanine, pentatonic acid, dipeptides and sphingolipids and terpenoids. The analysis also showed that adenosine was the most dominant non-volatile compound in the two types of rice bran cultivars.

5 In vivo Study of Fermented Rice Bran

The parameters that are widely tested in the *in vivo* study of fermented rice bran are total cholesterol(TC), triglycerides(TG), aspartate aminotransferase(AST), alanine aminotransferase(ALT), serum blood sugar levels and blood pressure. Fermented rice bran is a good source of antioxidants owing to its bioactive compounds such as phenolic compounds, flavonoids, carotenoids and anthocyanins. Many of these bioactive compounds are known to be useful as functional ingredients to prevent various chronic diseases.

The studies *in vivo* on fermented rice bran are presented in **Table 5**. The hepatoprotective effect on the administration of 0.4% w/w fermented rice bran orally in induced mice carbon tetrachloride $(CCl_4)^{58}$. The results of these studies indicate that fermented rice bran can prevent the incidence of liver damage caused by CCl_4 by increasing the antioxidant activity and decreasing AST and ALT. Another study showed the same results, i.e., a decrease in AST and ALT in the group of mice given fermented rice bran⁵⁹.

AST has been widely used as a marker to measure liver damage⁵⁸⁾. TNF- α , IL-6 and IL-1 β are pro-inflammatory cytokines that can cause colonic tissue damage and ulceration of the large intestine if present in excessive amounts⁶⁰⁾. The supplementation of fermented rice bran can protect the C57BL/6N mice from ulcerative colitis by decreasing TNF- α , IL-6 and IL-1 β , increasing the body weight and stool consistency and reducing intestinal bleeding due to a rise in short-chain fatty acids (SCFA) and tryptamine⁵⁹⁾. Another benefit of fermented rice bran is its anti-inflammatory effect which is achieved by reducing pro-inflammatory cytokines. The ovalbumin (OVA) model showed that fermented rice bran extract reduced TNF- α , interferon (IFN- γ), IL-6 and IL-10 in mice⁶¹⁾. Another study states that fermented rice bran extract can prevent atopic dermatitis by reducing T cells CD8⁺ and cells Gr-1⁺/CD11b⁺ B and inhibiting the expression of cytokine mRNA(IL-5 and IL-13) $^{62)}$.

Fermented rice bran has been reported to lower blood pressure when administered to stroke-prone spontaneously hypertensive rats (SHRSP)^{28, 38, 57)}. Both studies reported that the oral administration of fermented rice bran can reduce systolic blood pressure (sBP), improve blood sugar levels, reduce insulin resistance and increase nitric oxide (NO) in the blood^{28, 57)}. Furthermore, fermented rice bran can effectively lower blood pressure due to an increase in the inhibitory activity of the angiotensin-converting enzyme (ACE) in serum³⁸.

TC, TG, LDL cholesterol and high-density lipoprotein (HDL) cholesterol are an important part of the lipid fraction in the human body. The anti-hypercholesterolemic effect of fermented rice bran has also been reported⁶³⁾. Rice bran compounds such as γ -oryzanol can lower the cholesterol levels in the blood, lower the LDL cholesterol, and increase the HDL cholesterol. Mice orally administrated with fermented rice bran showed a decrease in TC, TG and LDL and increased blood HDL cholesterol^{38, 59, 63)}. Several possible mechanisms can occur in the improvement of lipid fractions in the blood, such as a decrease in lipid and cholesterol absorption as well as cholesterol synthesis and an increase in cholesterol secretion, HDL synthesis or antioxidant activity⁶³⁾.

6 Conclusions and Perspectives

Fermentation using the SSF technique is an alternative method that can be used to increase the number of active compounds in rice bran as a by-product of the rice milling process. The biochemical reactions that occur during the fermentation process lead to changes in the nature and characteristics of the rice bran. SSF can increase the TPC of rice bran. The volatile and non-volatile compounds present in the rice bran contribute to its aroma and flavour,

Fermented Rice Bran

Rice bran	Starter (microbes)	Animals	Treatment	Results	References
Fermented rice bran (Inpari 30)	Rhizopus oligosporus	SHRSP/ Izumo, male	40 mg/kg body weight	Systolic blood pressure, blood glucose, and HOMA index ↓ Plasma NO ↑	28)
Fermented rice bran	Aspergillus kawachii and Lactobacillus brevis, Lactobacillus rhamnosus, and Enterococcus faecium	SHRSP/ Izumo, male	2 g/kg body weight	Systolic blood pressure, diastolic blood, blood glucose, TC, LDL-C, and HOMA index ↓ ACE inhibition ↑ Serum TG ↓ p-AMPK, p-AMPKα, and HDL-C ↑	38)
Fermented rice bran	Bacillus sp., Bacillus subtilis, Bacillus sonolensis, Bacillus sirculans	ICR mice, male	0.4% body weight	Serum ALT, AST, ALP, and TC ↓ Serum TG ↑ Serum LDH ↓	58)
Fermented rice bran	Bacillus subtilis	Sprague Dawley rats, male	1.5% and 3% body weight	ALT, AST, and ALP ↓ Serum TG ↓ Liver TG ↓	59)
Fermented rice bran	Aspergillus kawachii and Lactobacillus brevis, Lactobacillus rhamnosus, and Enterococcus faecium	C57B1/6N mice, male	10% body weight	Intestine myeloperoxidase, intestine TBARS, TNF- α , IL-1 β , and IL-6 \downarrow Short chain fatty acid and tryptamine \uparrow	60)
Fermented rice bran extract	Issatchenkia orientalis	BALB/c mice, female	1.5 and 3 g/kg body weight	TNF- α , IL-10, and IL-6 \downarrow IFN- $\gamma \downarrow$	61)
Fermented rice bran	Lactobacillus rhamnosus and Pichia deserticola	NC/Nga mice, male	300 mg/kg body weight	IgE \downarrow CD4+, CD8+, and Gr-1/CD11b+ \downarrow	62)
Fermented rice bran extract	Rhizopus oligosporus	Sprague Dawley rats, male	1102.5 and 2205 mg/kg BW/day	Serum glucose, TC, TG, and serum LDL-C ↓ Serum HDL-C ↑	63)

Table 5In vivo studies of fermented rice bran.

Note: \uparrow = increasing; \downarrow = decreasing

which are formed during the fermentation process, and can improve the sensory quality of the rice bran. The increase in the number of active compounds in the rice bran is accompanied by a rise in bioactivity, which has been proven in the *in vivo* studies.

SSF is a simple technique for producing bioactive compounds in rice bran. This technique is economical because the materials used are agro-industrial by-products. Additionally, this technique is environment-friendly since it reduces the number of industrial by-products disposed into the environment. SSF can be used as an alternative method to improve the functional properties of rice bran as a food ingredient for health promotion in the future.

Acknowledgments

This work financially was supported by the Ministry of Research and Technology, Indonesia. The partial of the research was supported by a grant from the collaboration research of JSPS and Directorate Higher Education, Ministry of Education, Indonesia.

References

- Charaborty, M.; Pooja, S.B.; Vinod. 2018. Nutritional and therapeutic value of rice bran. *Inter. J. Green Herb. Chem.* 7, 451-461 (2018).
- 2) FAO. Rice market monitor. Vol. XXI(1). www.fao.org/ economic/RMM(accessed on 7 May 2021).
- Badan Pusat Statistik (BPS). Luas Panen dan Produksi Padi di Indonesia. Available online: https://www.bps.

go.id/pressrelease/2020/02/04/1752/luas-panen-danproduksi-padi-pada-tahun-2019-mengalami-penurunan-dibandingkan-tahun-2018-masing-masing-sebesar-6-15-dan-7-76-persen.html(accessed on 7 May 2021).

- 4) Henderson, A.J.; Ollila, C.A.; Kumar, A.; Borresen, E.C.; Raina, K.; Agarwal, R. Chemopreventive properties of dietary rice bran: Current status and future prospects. *Adv. Nutr. J.* **3**, 643-653 (2012).
- 5) Ryan, E.P. Bioactive food components and health properties of rice bran. J. Am. Vet. Med. Assoc. 238, 593-600(2011).
- 6) Li, W.; Chen, Z.; Yan, M.; He, P.; Chen, Z.; Dai, H. The protective role of isorhamnetin on human brain microvascular endothelial cells from cytotoxicity induced by methylglyoxal and oxygen-glucose deprivation. *J. Neurochem.* **136**, 651-659 (2016).
- 7) Aguilar-Garcia, C.; Gavino, G.; Baragano-Mosqueda, M.; Hevia, P.; Gavino, V.C. Correlation of tocopherol, tocotrienol, c-oryzanol and total polyphenol content in rice bran with different antioxidant capacity assays. *Food Chem.* **102**, 1228-1232 (2007).
- 8) Shin, H.Y.; Kim, S.M.; Lee, J.H.; Lim, S.T. Solid-state fermentation of black rice bran with *Aspergillus awamori* and *Aspergillus oryzae*: Effects on phenolic acid and antioxidant activity of bran extract. *Food Chem.* **272**, 235-241 (2019).
- 9) Perez-Ternero, C.; Werner, C.M.; Nickel, A.G.; Herrera, M.D.; Motilva, M.J. *et al.* Ferulic acid, a bioactive component of rice bran, improves oxidative stress and mitochondrial biogenesis and dynamics in mice and in human mononuclear cells. *J. Nutr. Biochem.* 48, 51-61 (2017).
- 10) Islam, Md.S.; Nagasaka, R.; Ohara, K.; Hosoya, T.; Ozaki, H. *et al.* Biological abilities of rice bran-derived antioxidant phytochemicals for medical therapy. *Curr. Topics Med. Chem.* **11**, 1847-1853 (2011).
- Cheng, J.; Choi, B.K.; Yang, S.H.; Suh, J.W. Effect of fermentation on the antioxidant activity of rice bran by *Monascus pilosus* KCCM60084. J. Appl. Biol. Chem. 59, 57-62 (2016).
- 12) Warren, B.E.; Farrell, D.J. The nutritive value of full fat and defatted Australian rice bran and chemical composition. *Anim. Feed Sci. Tech.* 27, 219-223 (1990).
- 13) Schmidt, C.G.; Goncalves, L.M.; Prietto, L.; Hackbart, H.S.; Furlong, E.B. Antioxidant activity and enzyme inhibition of phenolic acids from fermented rice bran with fungus *Rhizopus oryzae*. Food Chem. 146, 371-377 (2014).
- 14) Martins, S.; Mussatto, S.I.; Martínez-Avila, G.; Montanez-Saenz, J.; Aguilar, C.N.; Teixeira, J.A. Bioactive phenolic compounds: Production and extraction by solid-state fermentation. A review. *Biotechnol. Adv.*

29, 365-373 (2011).

- 15) Oliveira, M.S.; Feddern, V.; Kupski, L.; Cipolatti, E.P.; Badiale-Furlong, E.; de Souza-Soare, L.A. Changes in lipid, fatty acids and phospholipids composition of whole rice bran after solid-state fungal fermentation. *Bioresour. Technol.* **102**, 8335-8338(2011).
- 16) Srivastava, R.K. Enhanced shelf life with improved food quality from fermentation processes. J. Food Technol. Preserv. 2, 8-14(2018).
- 17) Oliveira, M.S.; Feddern, V.; Kupski, L.; Cipolatti, E.P.; Badiale-Furlong, E.; Souza-Soares, L.A. Physico-chemical characterization of fermented rice bran biomasscaracterización fisico-química de la biomasa del salvado de arroz fermentado. *CyTA-J. Food* 8, 229-236 (2010).
- 18) Cabuk, B.; Nosworthy, M.G.; Stone, A.K.; Korber, D.R.; Tanaka, T.; House, J.D.; Nickerson, M.T. Effect of fermentation on the protein digestibility and levels of non-nutritive compounds of pea protein concentrate. *Food Tech. Biotech.* 56, 257-264 (2018).
- Oliveira, M.D.S.; Cipolatti, E.P.; Furlong, E.B.; Soares, L.S. Phenolic compounds and antioxidant activity in fermented rice (*Oryza sativa*) bran. *Ciênc. Tecnol. Aliment. Campinas.* 32, 531-537 (2012).
- 20) Ardiansyah; Ariffa, F.; Astuti, R.M.; David; Handoko, D.D.; Budijanto, S.; Shirakawa, S. Non-volatile compounds and blood pressure-lowering activity of Inpari 30 and Cempo Ireng fermented and non-fermented rice bran. *AIMS Agric. Food* 6, 337-359 (2021).
- 21) Kupski, L.; Cipolatti, E.; Rocha, Md.; Oliveira, Md.S.; Furlong, E.B. Solid-state fermentation for the enrichment and extraction of proteins and antioxidant compounds in rice bran by *Rhizopus oryzae. Braz. Arch. Biol. Technol.* 5, 937-942 (2012).
- 22) Jamaluddin, A.; Razak, D.L.A.; Rashid, N.Y.A.; Sharifudin, S.A.; Kahar, A.A. *et al.* Effects of solid state fermentation by *Monascus purpureus* on phenolic content and biological activities of coconut testa and rice bran. *Adv. Nat. Prod. Res. Bioremed.* **78**, 23-28 (2016).
- 23) Razak, D.L.A.; Rashid, N.Y.A.; Jamaluddin, A.; Sharifudin, S.A.; Long, K. Enhancement of phenolic acid content and anti-oxidant activity of rice bran fermented with *Rhizopus oligosporus* and *Monascus purpureus. Biocat. Agric. Biotech.* 4, 33-38 (2015).
- 24) Chen, Y.; Ma, Y.; Dong, L.; Jia, X.; Liu, L. *et al.* Extrusion and fungal fermentation change the profile and antioxidant activity of free and bound phenolics in rice bran together with the phenolic bioaccessibility. *LWT Food Sci. Technol.* **115**, 108461 (2019).
- 25) Janarny, G.; Ghunatilake, K.D.P.P. 2020. Changes in rice bran bioactives, their bioactivity, bioaccessibility and bioavailability with solid-state fermentation by *Rhizopus oryzae. Biocatal. Agric. Biotechnol.* 23,

101510(2020).

- 26) Zulfafamy, K.E.; Ardiansyah; Budijanto, S. Antioxidative properties and cytotoxic activity against colon cancer cell WiDr of *Rhizopus oryzae* and *Rhizopus oligosporus*-fermented black rice bran extract. *Currt. Res. Nutr. Food Sci.* 6, 23-34 (2018).
- 27) Noviasari, S.; Kusnandar, F.; Setiyono, A.; Budi, F.S.; Budijanto, S. Profile of phenolic compounds, DPPHscavenging and anti α-amylase activity of black rice bran fermented with *Rhizopus oligosporus*. *Pertanika J. Trop. Agric. Sci.* **42**, 489-501 (2019).
- 28) Ardiansyah; David, W.; Handoko, D.D.; Kusbiantoro, B.; Budijanto, S.; Shirakawa, H. Fermented rice bran extract improves blood pressure and glucose in strokeprone spontaneously hypertensive rats. *Nutr. Food Sci.* 49, 844-853 (2019).
- 29) Rashid, N.Y.A.; Razak, D.L.A.; Jamaluddin, A.; Sharifuddin, S.A.; Long, K. Bioactive compounds and antioxidant activity of rice bran fermented with lactic acid bacteria. *Malaysian J. Microbiol.* **11**, 156-162 (2015).
- 30) Omarini, A.B.; Labuckas, D.; Zunino, M.P.; Pizzolitto, R.; Fernandez-Lahore, M. *et al.* Upgrading the nutritional value of rice bran by solid-state fermentation with *Pleurotus sapidus. Fermentation* 5, 44 (2019).
- 31) Kim, D.; Dong Han, G. Fermented rice bran attenuates oxidative stress. in Wheat and rice in disease prevention and health: Benefits, risks and mechanisms of whole grains in health promotion (Watson, R.D.; Preedy, V.R.; Zibadi, S. eds.). Academic Press, Elsevier, pp. 467-480 (2014).
- 32) Razak, D.L.; Jamaluddin, A.; Sharifudin, S.A.; Kahar, A.A.; Long, K. Cosmeceutical potentials and bioactive compounds of rice bran fermented with single and mix culture of *Aspergillus oryzae* and *Rhizopus oryzae*. J. Saudi Soc. Agric. Sci. 16, 127-134 (2017).
- 33) Kurniawati, T.; Indrati, R.; Sardjono. Isolation of *Rhi-zopus oryzae* from rotten fruit and its potency for lactic acid production in glucose medium with and without addition of calcium carbonate. *Agritech.* 34, 170-176 (2014).
- 34) Denardi-Souzaa, T.; Massaroloa, K.C.; Tralamazzab, S.M.; Badiale-Furlonga, E. Monitoring of fungal biomass changed by *Rhizopus oryzae* in relation to amino acid and essential fatty acids profile in soybean meal, wheat and rice. *CyTA J. Food* 16, 156-164 (2018).
- 35) Ribeiro, A.C.; Graça, C.D.; Chiattoni, L.M.; Massarolo, K.C.; Duarte, F.A. *et al.* Fermentation process in the availability of nutrients in rice bran. *Res. Rev. J. Microbiol. Biotechnol.* 6, 45-52 (2017).
- 36) Hernandez, L.L.; Bolívar, G.; Toro, C.R. Effect of solid state fermentation with *Rhizopus oryzae* on biochemical and structural characteristics of sorghum (*Sorghum bicolor*(L.)Moench). *Inter. Food Fer-*

ment. Technol. 8, 27-36 (2018).

- 37) Hartanti, A.T.; Rahayu, G.; Hidayat, I. *Rhizopus* species from fresh tempeh collected from several regions in Indonesia. *HAYATI J. Biosci.* **22**, 136-142 (2015).
- 38) Alauddin, Md.; Shirakawa, H.; Koseki, T.; Kijima, N.; Ardiansyah *et al.* Fermented rice bran supplementation mitigates metabolic syndrome in stroke-prone spontaneously hypertensive rats. *BMC Complem. Alternat. Med.* 16, 442 (2016).
- 39) Bhanja, T.; Rout, S.; Banerjee, R.; Bhattacharyya, B.C. Studies on the performance of a new bioreactor for improving antioxidant potential of rice. *LWT Food Sci. Tech.* **41**, 1459-1465 (2019).
- 40) Mitsuda, H.; Yasumoto, K.; Iwami, K. Analysis of volatile components in rice bran. Agric. Biol. Chem. 32, 453-58(1968).
- 41) Tsugita, T.; Kurata, T.; Fujimaki, M. Volatile components in the steam distillate of rice bran: Identification of neutral and basic compounds. *Agric. Biol. Chem.* 42, 643-651 (1978).
- 42) Zeng, M.; Zhang, L.; He, Z.; Qin, F.; Tang, X. et al. Determination of flavor components of rice bran by GC-MS and chemometrics. Food Anal. Meth. 4, 539-545 (2012).
- 43) Sukhontara, S.; Theerakulkait, C.; Miyazawa, M. Characterization of volatile aroma compounds from red and balck rice bran. *J. Oleo Sci.* **58**, 155-161 (2009).
- 44) Jordan, M.J.; Margaria, C.A.; Shaw, P.E.; Goodner, K.L. Volatile components and aroma active compounds in aqueous essence and fresh pink guava fruit puree (*Psidium guajava* L.) by GC-MS and multidimensional GC/GC-O. J. Agric. Food Chem. 51, 1421-1426 (2003).
- 45) Mosciano, G. Nonanoic acid(pelargonic acid). The Good Scents Company: http://www.thegoodscentscompany.com/data/rw1012131.html(accessed on 1 May 2021).
- 46) Luebke, W. Hexahydrofarnesyl acetone (6,10,14-trimethyl-2-pentadecanone). The Good Scents Company: http://www.thegoodscentscompany.com/data/ rw1309781.html(accessed on 1 May 2021).
- 47) Yang, D.S.; Shewfelt, R.L.; Lee, K.S.; Kays, S.J. Comparison of odor-active compounds from six distinctly different rice flavor types. J. Agric. Food Chem. 56, 2780-2787 (2008).
- 48) Luebke, W. Hexanoic acid(Caproic acid). The Good Scents Company: http://www.thegoodscentscompany. com/data/rw1008541.html(accessed on 1 May 2021).
- 49) Mosciano, G. Myristic Acid(tetradecanoic acid). The Good Scents Company: http://www.thegoodscentscompany.com/data/rw1009061.html(accessed on 1 May 2021).
- 50) Zarei, I.; Brown, D.G.; Nealon, N.J.; Ryan, E.P. Rice bran metabolome contains amino acids, vitamins & co-

factors, and phytochemicals with medicinal and nutritional properties. *Rice* 10, 24(2017).

- 51) Zarei, I.; Luna, E.; Leach, J.E.; McClung, A.; Vilchez, S. et al. Comparative rice bran metabolomics across diverse cultivars and functional rice gene-bran metabolite relationships. *Metabolites* 8, 63 (2018).
- 52) Suttiarporn, P.; Chumpolsri, W.; Mahatheeranont, S.; Luangkamin, S.; Teepsawang, S.; Leardkamolkarn, V. Structures of phytosterols and triterpenoids with potential anti-cancer activity in bran of black non-glutinous rice. *Nutrients* 7, 1672-1687 (2015).
- 53) Kim, S.K.; Van, T.Q. Potential beneficial effects of marine algal sterols on human health. Adv. Food Nutr. Res. 64, 191-198(2011).
- 54) Somintara, S.; Leardkamolkarn, V.; Suttiarporn, P.; Mahatheeranont, S. Anti-tumor and immune enhancing activities of rice bran gramisterol on acute myelogenous leukemia. *PLoS One* **11**, 1-19 (2016).
- 55) Gabayy, O.; Sanchez, C.; Salvat, C.; Chevy, F.; Breton, M. et al. Stigmasterol: A phytosterol with potential anti-osteoarthritic properties. Ostheoarthitis Cartilage 18, 106-116 (2010).
- 56) Ogbe, R.J.; Ochalefu, D.O.; Mafulul, S.G.; Olaniru, O.B. A review on dietary phytosterols: Their occurrence, metabolism and health benefits. *Asian J. Plant Sci. Res.* 5, 10-21 (2015).
- 57) Ardiansyah; Nada, A.; Rahmawati, N.T.I.; Oktriani, A.; David, W. *et al.* Volatile compounds, sensory profile and phenolic compounds in fermented rice bran. *Plants* **10**, 1073 (2021).
- 58) Park, S.L.; Lee, S.Y.; Nam, Y.D.; Yi, S.H.; Seo, M.J.; Lim, S.I. Hepatoprotective effect of fermented rice bran

against carbon tetrachloride-induced toxicity in mice. *Food Sci. Biotechnol.* **23**, 165-171 (2014).

- 59) Ahn, H.Y.; Cho, Y.S. An animal study to compare hepatoprotective effects between fermented rice bran and fermented rice germ and soybean in a sprague-dawley rat model of alcohol-induced hepatic injury. *Multidicipl. Sci. J.* **3**, 54-66 (2020).
- 60) Islam, J.; Koseki, T.; Watanabe, K.; Ardiansyah; Budijanto, S. *et al.* Dietary supplementation of fermented rice bran effectively alleviates dextran sodium sulfateinduced colitis in mice. *Nutrients* 9, 747 (2017).
- 61) Fan, J.P.; Choi, K.M.; Han, G.D. Inhibitory effects of water extracts of fermented rice bran on allergic response. *Food Sci. Biotechnol.* **19**, 1573-1578(2010).
- 62) Saba, E.; Lee, C.H.; Jeong, D.H.; Lee, K.; Kim, T.H. *et al.* Fermented rice bran prevents atopic dermatitis in DNCB-treated NC/Nga mice. *J. Biomed Res.* **30**, 334-343(2016).
- 63) Nurrahma, B.A.; Suryajayanti, M.F.; Dewi, A.L.; Khairia, Z.; Kusuma, R.J.; Suyoto. Fermented rice bran extract improves dyslipidemia in rodents. *Nutr. Food Sci.* 48, 375-383 (2018).

CC BY-SA 4.0 (Attribution-ShareAlike 4.0 International). This license allows users to share and adapt an article, even commercially, as long as appropriate credit is given and the distribution of derivative works is under the same license as the original. That is, this license lets others copy, distribute, modify and reproduce the Article, provided the original source and Authors are credited under the same license as the original.

